



## Enhanced selective metal adsorption on optimised agroforestry waste mixtures



Emilio Rosales<sup>a,b,\*</sup>, Laura Ferreira<sup>a</sup>, M. Ángeles Sanromán<sup>a</sup>, Teresa Tavares<sup>b</sup>, Marta Pazos<sup>a</sup>

<sup>a</sup> Department of Chemical Engineering, University of Vigo, Isaac Newton Building, Campus As Lagoas Marcosende, 36310 Vigo, Spain

<sup>b</sup> Centro de Engenharia Biológica, Universidade do Minho, Campus de Gualtar, 4710-057 Braga, Portugal

### HIGHLIGHTS

- Agroforestry wastes present good sorption capacities for a mixture of several metals.
- A mixture of biosorbents increases the total removal of metals.
- Optimisation of the biosorbent mixture proportions maximises metal removal.
- Recovery of metals can be achieved by treatment with weak acids.

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### ABSTRACT

The aim of this work is to ascertain the potentials of different agroforestry wastes to be used as biosorbents in the removal of a mixture of heavy metals. Fern (FE), rice husk (RI) and oak leaves (OA) presented the best removal percentages for Cu(II) and Ni(II), Mn(II) and Zn(II) and Cr(VI), respectively. The performance of a mixture of these three biosorbents was evaluated, and an improvement of 10% in the overall removal was obtained (19.25 mg/g). The optimum mixture proportions were determined using simplex-centroid mixture design method (FE:OA:RI = 50:13.7:36.3). The adsorption kinetics and isotherms of the optimised mixture were fit by the pseudo-first order kinetic model and Langmuir isotherm. The adsorption mechanism was studied, and the effects of the carboxylic, hydroxyl and phenolic groups on metal–biomass binding were demonstrated. Finally, the recoveries of the metals using biomass were investigated, and cationic metal recoveries of 100% were achieved when acidic solutions were used.

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## 1. Introduction

The processes of a wide variety of industries such as tanneries, plating, textile, mining, cosmetics, pharmaceutical and paper use many metals ions, including Cr(VI), Cr(III), Mn, Cu, Zn and Ni (Gupta et al., 2009). Once used, the compounds containing these metals can generate a wide range of wastewaters containing different pollutants. The amount of metal present in the wastewater is related to the operations developed in the industry. The removal of metals in effluent is required to attain the safe permissible metal limits and avoid the toxic and poisonous effects of these metals in

the environment. For example, Cr(VI) has been recognised as a carcinogenic and mutagenic agent (Brauer and Wetterhahn, 1991), and the presence of Cr(III) at concentrations exceeding 5.0 mg/L may be toxic for aquatic life (Pérez Marín et al., 2009).

A wide range of physical–chemical, biological and oxidative technologies have been proposed to treat wastewaters containing metals (Nguyen et al., 2013). Among the different treatments available, adsorption is one of the most attractive technologies because of its versatility, simple design, ease of operation and high efficiency for pollutant removal (Rosales et al., 2012). However, the main drawback of adsorption is the high cost of efficient adsorbent materials. Activated carbon is the most widely studied adsorbent and has been widely applied in industry, providing high removal rates; however, its cost is a drawback in its general use (Crini, 2006). Thus, alternative low cost and biodegradable adsorbents are required. A suitable low-cost adsorbent requires a porous structure, mechanical and chemical stability and affinity for the target pollutants (Cobas et al., 2014). Recent approaches suggest

\* Corresponding author at: Department of Chemical Engineering, University of Vigo, Isaac Newton Building, Campus As Lagoas Marcosende, 36310 Vigo, Spain. Tel.: +34 986 812 304; fax: +34 986 812 380.

E-mail addresses: [emiliov@uvigo.es](mailto:emiliov@uvigo.es) (E. Rosales), [ferreira@uvigo.es](mailto:ferreira@uvigo.es) (L. Ferreira), [sanroman@uvigo.es](mailto:sanroman@uvigo.es) (M. Ángeles Sanromán), [ttavares@deb.uminho.pt](mailto:ttavares@deb.uminho.pt) (T. Tavares), [mcurras@uvigo.es](mailto:mcurras@uvigo.es) (M. Pazos).

## Nomenclature

CA	cane pruning wine grapes	$K_S$	Sips model isotherm constant (L/g)
OA	oak leaves	$a_s$	Sips model constant (L/mg)
RI	rice husk	$\beta_S$	Sips model exponent
GR	grapefruit peelings	$k_1$	Pseudo-first order rate constant (1/min)
FE	fern	$k_2$	Pseudo-second rate constant (g/(mg min))
ME	mixture fern, oak leaves and rice husk	$C_e$	concentration of metal solution at adsorption equilibrium (mg/L)
OAM	optimised biosorbent mixture	$C_i$	initial metal concentration of metal solution (mg/L)
$q_{max}$	model maximum achievable uptake by a system (mg/g)	$q_e$	equilibrium adsorption capacity of biosorbent (mg/g)
$b_L$	langmuir model affinity between the sorbate and sorbent (L/mg)	$q_t$	amount of metal adsorbed per unit of biosorbent at time $t$ (mg/g)
$R^2$	linear correlation coefficient	$t$	contact time (min)
$K_F$	Freundlich model binding capacity (mg/g)/(mg/L) <sup>1/n</sup>	$R$	universal gas constant (J/(mol g))
$n_F$	Freundlich model affinity between the sorbent and sorbate	$T$	temperature (°C)
$K_{RP}$	Redlich–Peterson model isotherm constant (L/g)	$k_{id}$	intra-particle diffusion rate constant (mg/(g h <sup>0.5</sup> ))
$a_{RP}$	Redlich–Peterson model constant (L/mg) <sup><math>\beta_{RP}</math></sup>		
$\beta_{RP}$	Redlich–Peterson model exponent		

the utilisation of non-conventional adsorbents; natural clays (Iglesias et al., 2013; Padilla-Ortega et al., 2013), various industrial and agricultural materials such as hazelnut straws, peanut shell, banana peel, orange peel, walnut shell, rice husk and sawdust (Nguyen et al., 2013; Alothman et al., 2013; Ma et al., 2014; Park et al., 2008; Krishnani et al., 2008; Abdolali et al., 2014a) and forestry materials such as fern or oak leaves (Prasad and Freitas, 2000; Barriada et al., 2009) have been tested. These agroforestry materials have been mainly applied to the removal of a single or a couple of metals, while little attention has been paid to the simultaneous adsorption of multiple metals (Krishnani et al., 2008; Abdolali et al., 2014b; Hossain et al., 2014a).

The goal of this study is to ascertain the abilities of different agroforestry wastes as alternative low-cost biosorbents for the removal of wastewater streams containing different metals. First, the adsorption potentials of the agroforestry wastes were investigated. Then, the performance of the combination of the biosorbents were evaluated, and the composition of the mixed biosorbents was optimised using response surface methodology. Isotherms and kinetic data were also obtained. Finally, the adsorption mechanism was studied, and biosorbent recovery was evaluated.

## 2. Experimental

### 2.1. Reagents

A stock solution containing Cr, Cu, Mn, Ni and Zn was obtained by dissolving the metal salts in water. ZnCl<sub>2</sub>, CuCl<sub>2</sub>·2H<sub>2</sub>O, MnCl<sub>2</sub>·4H<sub>2</sub>O and NiCl<sub>2</sub>·6H<sub>2</sub>O were purchased from Merck, and potassium dichromate (K<sub>2</sub>Cr<sub>2</sub>O<sub>7</sub>) was obtained from Sigma–Aldrich. All the used chemicals were reagent grade.

### 2.2. Agroforestry wastes

The materials used in the study were agriculture and forestry wastes. Cane pruning wine grapes (CA), fern (FE), eucalyptus bark (EU), oak leaves (OA), grapefruit (GR) and rice husk (RI) were provided by local suppliers. The wastes were washed three times with distilled water, dried in an oven at 60 °C for 24 h and then grounded to pass through a 1-mm sieve before use.

### 2.3. Metal adsorption by biosorbents

Adsorption studies were performed individually for each biosorbent in Erlenmeyer flasks (250 mL) containing 1 g of biosorbent with a working volume of 150 mL. The experiments were carried out at 25 °C and 150 rpm for 4 h. A synthetic wastewater was prepared containing a mixture of the different metals. The initial concentration of each metal in this synthetic wastewater was 50 mg/L, resulting in a total metal content of 250 mg/L.

The metal adsorption assays using a mixture of untreated biosorbents were performed under the same conditions with a final mass of 1 g. Kinetic and isotherm studies were carried out under the same conditions as previously described. For the isotherms assays, the concentration of each metal in the synthetic wastewater ranged from 6–400 mg/L.

### 2.4. Adsorption process mechanism

The adsorption mechanism was evaluated by modifying the surfaces of the biosorbents using different selected chemical agents. Chemical modification methods could activate/deactivate the available binding sites on the biomass surface (Abdolali et al., 2014a). To evaluate the adsorption mechanism, the chemical blocking of functional groups was applied. The esterification of carboxylic acids with methanol-HCl (HM-ME) and the methylation of hydroxyl and phenolic groups with formaldehyde (FO-ME) were carried out. The carboxylic acids present in the biosorbents were esterified by alcohols in the presence of a suitable acidic catalyst (HCl) according to Gardea-Torresdey (1990). The modification of the biosorbents with formaldehyde was performed according to Deshkar et al. (1990).

### 2.5. Recovery of the metals

After adsorption, the biosorbent was recovered by filtration and dried in a stove at 60 °C for 24 h. The biosorbent loaded with metals was then treated with an eluent (water, HCl 0.1 N and HNO<sub>3</sub> 0.1 N). Desorption tests were performed using 1 g of the used biosorbent in 150 mL of the eluent. After 120 min at 150 rpm, the samples were filtered, and the pH and metal concentrations in the liquid were determined.

## 2.6. Optimisation of biosorbent mixture

To optimise the mixture, a tool-like experimental design was selected. This design allows a reduction in the number of experiments required to optimise the biosorbent composition and to fit a model equation to the obtained data.

The optimum composition of the biosorbent mixture was determined using simplex-centroid mixture design (SCMD). SCMD is a type of experimental design used to analyse the response of two or more components to obtain a final product with a different composition. The proportion  $X_i$  of a component in the mixture must satisfy the constraints  $0 \leq X_i \leq 1$ , where  $i = 1, 2, \dots, q$ , and  $\sum X_i = 1$ , where  $q$  is the number of components. The geometric description of the spatial representation will depend on the number of components  $q$  (a line for  $q = 2$ , an equilateral triangle for  $q = 3$ , etc.) (Yoshiara et al., 2012).

The regression model equations were fitted to a quadratic expression as follows:

$$Y_i = \sum b_j X_j + \sum b_{jk} X_j X_k \quad (1)$$

where  $Y$  is the response,  $b$  are the constants and  $X$  are the independent parameters. Statistical analysis of the model was performed to evaluate the analysis of variance (ANOVA) using Design Expert® 8.0.0 software (Stat-Ease Inc., Minneapolis, USA).

## 2.7. Determination procedures

### 2.7.1. Metals determination

The metal concentrations in the supernatant were determined using ICP-OES (PerkinElmer-Optima 8000). Hexavalent chromium was quantified by measuring the absorbance at 540 nm of the purple complex of Cr(VI) with 1,5-diphenylcarbazide in acidic solution. The Cr(III) concentration was calculated as the difference between the total chromium and the Cr(VI) concentration.

### 2.7.2. Adsorption uptake and removal percentages

The amount of metal adsorbed by the biosorbent (uptake) was calculated using the following expression:

$$q_t = (C_i - C_t) \cdot V/M \quad (2)$$

where  $C_i$  and  $C_t$  (mg/L) are the solute concentration at the beginning and through time ( $t$ ) in the solution, respectively,  $q_t$  is the solute uptake (mg/g),  $V$  is the volume of the solution (L) and  $M$  is the mass of the biosorbent (g).

The removal percentage, (%), was calculated using Eq. (3):

$$\text{Metal removal} = 100 \cdot (C_i - C_t)/C_i \quad (3)$$

where  $C_i$  and  $C_t$  (mg/L) are the solute concentrations at the beginning and through time ( $t$ ) in the solution, respectively.

The obtained concentration and uptake profiles were fitted by a suitable kinetic or isotherm equation, and the rate constants or adsorption parameters were calculated using Sigma Plot 4.00 software. The Sigma Plot curve fitter uses an iterative procedure based on the Marquardt–Levenberg algorithm, which seeks the values of the parameters that minimise the sum of the squared differences between the observed and predicted values of the dependent variable.

### 2.7.3. Characterisation of biosorbents

The zero point charge (ZPC) was determined by mass titration (Noh and Schwarz, 2013). Approximately 50 mL of 0.1 M  $\text{NaNO}_3$  as the background electrolyte was transferred into a 250 mL of Erlenmeyer flask, and the solution pH was adjusted with diluted HCl and NaOH to the desired values (3 and 11). The final volume of the solution was 50 mL. The mixture was shaken for 24 h and

then filtered through filter paper. The pH in filtrate was measured with a CyberScanpH2100 Bench Meter (Eutech Instruments) after three-point calibration.

The acidic surface functional groups were quantified by Boehm titration (Boehm, 1966). First, a 0.3 g sample was mixed with 50 mL of 0.05 mol/L NaOH solution in a 250 mL Erlenmeyer flask. The flask was sealed and shaken for 24 h at room temperature. Then, the sample was separated by filtration, and the filtrate was titrated with 0.05 mol/L of HCl standard solution. Finally, the amounts of the functional groups per gram of waste were calculated according to the hypothesis that carboxylic, lactonic and phenolic groups can react with NaOH. The identification of basic groups (pirone and carbanion) was analogous to Boehm titration using HCl as acid.

Fourier-transform infrared (FT-IR) spectra of the biosorbents were recorded on an FT-IR spectrometer (model FT-IR/4100, Jasco). The samples were previously dried in an oven at 60 °C for 40 min.

Scanning electron microscopy (SEM) was performed on a JEOL JSM-6700F equipped with an EDS (Oxford Inca Energy 300, Electron Microscopy Service, C.A.C.T.I., University of Vigo) using an accelerating voltage of 15 kV. The analysed biosorbents were coated with carbon for SEM observation.

## 3. Results and discussion

### 3.1. Characterisation of biosorbents

The chemical structure of the biosorbent is a key factor in the adsorption process; the selected biosorbents have been previously characterised using FTIR spectroscopy to determine the surface functional groups involved. The FTIR spectra are shown in the [Supplementary data \(S.1\)](#).

The selected agroforestry wastes present high carbohydrates/lignin content, suggesting the presence of functional groups (methoxyl, phenolic, hydroxyl, aliphatic hydroxyl, carbonyl, etc.). Some of these groups suggest that the metal binding takes place by chelation, as reported by Komy et al. (2013). The most representative bands detected were identified, and similar functional groups were found on the adsorbents. The bands in the 3450–3200  $\text{cm}^{-1}$  region could be indicative of the complex nature of the examined biomasses and assigned to hydroxyl and amine groups (Komy et al., 2013). The bands assigned to asymmetric and symmetric methyl and methylene stretching are located at 2950 and 2850  $\text{cm}^{-1}$ , carbonyl stretching corresponds to the band at 1800–1650  $\text{cm}^{-1}$ , the amide I and amide II bands are observed at 1652 and 1531  $\text{cm}^{-1}$  (Li et al., 2010) and the band at 1050–1020  $\text{cm}^{-1}$  is assigned to carbohydrate units (Barriada et al., 2009; Abdolali et al., 2014b; Komy et al., 2013). The hydroxyl and carboxyl functional groups are suitable for the adsorption and binding of heavy metals (Hossain et al., 2014b). These facts are in accordance with the reports of Barriada et al. (2009), Nguyen et al. (2013), who analysed agriculture and forestry wastes in the removal of metals. The FTIR spectra of the selected biosorbents verify that the biosorbents are mainly composed of lignocellulosic materials (lignin, cellulose and hemicellulose).

SEM pictures of some adsorbents (FE, OA, and RI) are shown in the [Fig. S3a](#). Different structures were detected in FE and homogeneous structures for OA and RI. The EDS analyses are shown in [Fig. S3b](#) and the elemental composition detected consisted only in carbon and oxygen in the surface of the selected agroforestry waste (FE and OA) but the RI presented a small amount of P, K and Mg.

The surface characteristics of the wastes can be determined using the point of zero charge (PZC). The charge of the surface may be positive, negative or metal, and this charge determines the favoured adsorption. At  $\text{pH} > \text{pH}_{\text{PZC}}$  the adsorption of cations

**Table 1**  
Removal percentage of metals by each agroforestry waste biosorbent.

	Metal removal (%)					
	Cu	Ni	Mn	Cr	Zn	pH
FE	94.50	40.48	11.84	64.60	41.54	4.52
OA	83.40	26.40	8.94	91.49	27.65	4.12
RI	68.77	21.77	50.50	6.33	89.74	4.83
CA	70.44	21.74	9.19	47.78	32.00	5.18
GR	49.76	14.62	6.61	0.26	24.89	4.97
ME	85.45	34.60	34.16	71.52	70.66	5.08

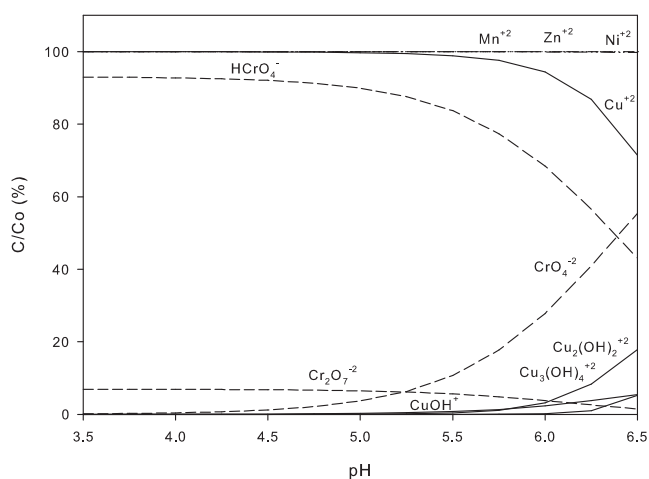
is favoured, while at  $\text{pH} < \text{pH}_{\text{PZC}}$  is favoured the adsorption of anions (Li et al., 2010). The  $\text{pH}_{\text{PZC}}$  values determined for FE, RI and OA were 5.33, 6.11 and 4.29, respectively. These results suggest that FE and OA have a strong cation adsorption ability. Meanwhile, FE and RI present higher capabilities for anion adsorption.

### 3.2. Metal speciation

The selected wastes were evaluated to remediate a synthetic wastewater containing a mixture of five metals: Cu(II), Cr(VI) Ni(II), Zn(II) and Mn(II). pH plays an important role in the adsorption process by affecting the surface charge of the biosorbent, the degree of ionisation and the speciation of the adsorbate (Yao et al., 2010). The experiments in this study were conducted at the natural pH of the multi-metal solution ( $\text{pH} \sim 5$ ); Table 1 shows the variation in the experimental pH at the end of the experiments with different biosorbents. Acidification of the wastewater was observed in the experiments with OA ( $\text{pH}$  4.1) and FE ( $\text{pH}$  4.53). Based on these results, the metal speciation in the synthetic wastewater should be considered. Metal speciation greatly determines the behaviour and toxicity of metals in the environment. The metal speciation was obtained using the modelling software Visual MINTEQ (Fig. 1). At the initial metal concentrations and the natural solution pH, Ni(II), Zn(II) and Mn(II) were present in their divalent cationic forms. Less than 1% of the present Cu existed as the monovalent cation ( $\text{CuOH}^+$ ), while Cr was present in three different anionic forms ( $\text{HCrO}_4^-$ ,  $\text{Cr}_2\text{O}_7^{2-}$  and  $\text{CrO}_4^{2-}$ ), with the monovalent anion  $\text{HCrO}_4^-$  being prominent. Considering the final pH observed in the different experiments, no changes in the metal speciation are expected.

### 3.3. Screening of biosorbents

The adsorption of ions present in the synthetic multi-metal solution by agroforestry wastes was evaluated.

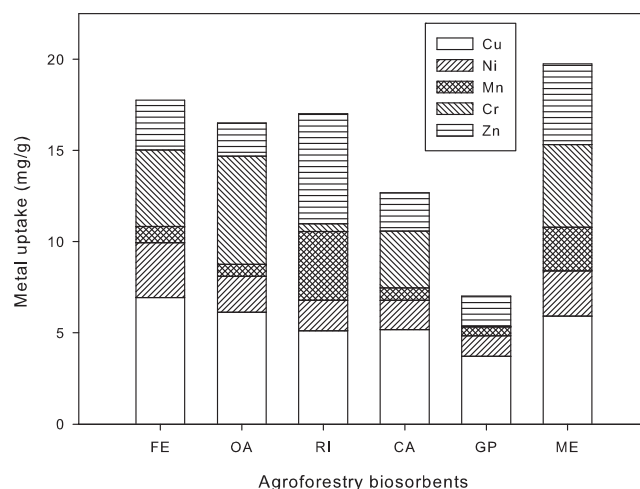


**Fig. 1.** Metal speciation of the mixture in the pH range of adsorption (data obtained based on software Visual MINTEQ).

The multi-metal mixture removal efficiencies of the five biosorbents are shown in Table 1. Among all metals, the maximum removals of Cr(VI), Zn(II) and Cu(II) were higher than those obtained for the other metals. The maximum removals achieved on the different biosorbents were nearly 90% for Cr(VI), Zn(II) and Cu(II). However, the obtained metal removal percentages for Mn(II) and Ni(II) were lower than 55% for all of the tested biosorbents. The highest removals of copper and nickel were obtained by FE; RI performed the best for manganese and zinc removal, and OA was optimal for chromium removal. These results are in accordance with those reported by Fontes and Gomes (2003) and can be explained by the high affinity of cations for the carboxylic and hydroxyl groups of cellulose. This aspect is reinforced by the previously obtained IR spectra of the biosorbents, which indicate the presence of these carboxylic and hydroxyl groups. One special consideration pertains to chromium because it is present as an anion; (Krishnani et al., 2008) and other authors have suggested that carboxylic and hydroxyl groups present in pretreated rice husk probably act as electron donors in the reduction of chromium to Cr(III) and subsequent adsorption.

The acidic and basic contents of the adsorbents are directly connected with the metal binding properties. Therefore, it is necessary to estimate the different types of binding to discuss the adsorption mechanism. The obtained results show the amount of binding points in acidic sites (FE 1.813 mmol/g, OA 2.917 mmol/g, RI 0.458 mmol/g, GR 1.167 mmol/g, EU 1.917 mmol/g) and basic sites (FE 3.208 mmol/g, OA 3.000 mmol/g, RI 3.167 mmol/g, GR 2.917 mmol/g, EU 3.000 mmol/g). The amount of acidic sites present in the different wastes reflects the heterogeneity of the source materials.

The selected materials have been shown to effectively bind a number of heavy metal ions, but it is unlikely that all ions were bound equally. Hossain et al. (2014b) suggested that the binding capacity for individual ions is different from a unique metal adsorption system, although the adsorption depends on a number of factors such as ionic potential, ionic radius, chemical properties and hydrolysis. Fig. 2 shows the total adsorbed metals and the distribution of different ions related to the total content. The total amounts of metals adsorbed using FE, RI and OA ranged from 16.51 to 17.71  $\text{mg}_{\text{metals}}/\text{g}_{\text{biosorbent}}$ . Based on the obtained values, a competitive adsorption order could be determined. The sequences were different for each biosorbent evaluated. For FE and OA, the general binding affinity for the studied metals decreasing in the following order: Cu(II) > Cr(VI) > Ni(II) > Zn(II) > Mn(II). Meanwhile, the order for RI was Zn(II) > Cu(II) > Mn(II) > Ni(II) > Cr(VI), that for CA was Cu(II) > Cr(VI) > Zn(II) > Ni(II) > Mn(II), and that of GP was Cu(II) > Zn(II) > Ni(II) > Mn(II) > Cr(VI).



**Fig. 2.** Metal uptake (mg/g) obtained for the different natural wastes: Cu, Ni, Mn, Cr and Zn.

**Table 2**

Analysis of functional groups in the mixture of biosorbents FE, OA, and RI in a ratio 1:1:1 for metal removal.

Wavelength range (cm <sup>-1</sup> )		Transmittance (%)			Bond/functional group
Frequency	Difference	Unloaded	Loaded	Difference	
3500–3200	2.8926	97.0729	99.2113	2.1384	OH stretches from hydroxyl, phenolic and carboxylic groups* (various polysaccharides, alcohols)
3000–2850	1.9285	92.4529	93.6298	1.1769	H–C–H stretch*, H–C–H asymmetric and symmetric stretch/alkanes** (various aliphatic compounds)
1820–1680	0.00	91.0512	91.9744	0.9232	C=O, stretching vibration of carboxylate functional group* (pectin)
1550–1475	-0.9642	93.5546	93.5558	0.0012	C–C stretching in aromatic skeletons (lignin) N–O asymmetric stretch**
1475–1450	0.00	90.9117	91.6175	0.7058	C–H band/alkanes** (lignin)
1450–1300	-1.9285	90.102	91.3539	-1.2519	Carbohydrate units (cellulose)
1300–1000	5.00	69.061	74.0612	4.8212	Carbohydrate units (cellulose/lignin/starch)

\* Ma et al., 2014.

\*\* Abdolali et al., 2014a.

All the selected biosorbents presented a high affinity for the adsorption of Cu(II). Cu is known to form complexes with very high stabilities despite the poor basicity of the donor group (Krishnani et al., 2008).

### 3.4. Biosorbent mixture adsorption and optimisation

#### 3.4.1. Biosorbent mixture

None of the selected biosorbents was able to remove all the metals in significant quantities. Therefore, the biosorbents that showed the best performances in metal removal were selected (FE–OA–RI), a mixture of these biosorbents was tested, and the results were compared with those obtained for the individual biosorbents. Initially, a 1:1:1 mixture of the three selected biosorbents was tested (ME). The attained results (Fig. 2) indicate an improvement in the total quantity of metals removed compared with the individual biosorbents because it is possible to remove a larger quantity of the combined metals using the mixture rather than using an individual biosorbent. The use of the mixture provides a selective environment for the adsorption of the different heavy metals (Barriada et al., 2009; Abdolali et al., 2014b). The improvement of total metal quantity removed using the mixture was more than 10% (19.25 mg/g of biosorbent), and the metals exhibited a more homogeneous affinity for ME in comparison with the individual biosorbents. Removal percentages higher than 70% were achieved for Cu(II), Cr(VI) and Zn(II), while removal percentages higher than 30% were obtained for Ni(II) and Mn(II) (Table 1).

FTIR analysis of the samples was carried out before and after the loading of the biosorbent mixture. The chemical shift and intensity of some bands changed due to heavy metal adsorption. The differences detected in the spectra are shown in Table 2. The band corresponding to the C–O stretching of carboxylic acid, esters and ethers (1300–1000 cm<sup>-1</sup>) shifted to higher values. The changes in the peaks in the range of 3500–3200 and 1820–1680 cm<sup>-1</sup> indicate the involvement of OH from hydroxyl, phenolic and carboxylic groups and the C=O from carboxylate functional groups, respectively (Barriada et al., 2009; Abdolali et al., 2014b; Yao et al., 2010; Popescu et al., 2007).

#### 3.4.2. Biosorbent mixture optimisation

The composition of the mixture of the three wastes was evaluated and optimised. An experimental design based on SCMD was selected.

A design of experiments was developed with a mixture of three components: FE, X<sub>1</sub>; OA, X<sub>2</sub>; and RI, X<sub>3</sub>. The selected response was the metal removal percentage, Y<sub>i</sub> (where i = Cu, Ni, Mn, Cr, Zn). The response to a mixture depends on the proportions of the components present, and a restriction related to the biosorbent proportions (X<sub>1</sub> + X<sub>2</sub> + X<sub>3</sub> = 1) was incorporated to the model. 14 runs were required to develop a model and the response for each metal removal (Table S1). The five obtained responses (metal removals) were transformed in polynomial equations in function of the pro-

portion of the three components (FE, OA, and RI) (Table S2). These equations can be plotted by contour graphs. The mixture contour plots for each response are shown in Fig. 3. The diagrams illustrate the effect of the biosorbent mixture for each metal. A high proportion of FE in the mixture is required to obtain high removals of Cu(II) and Ni(II). However, the removals of Mn(II) and Zn(II) increase when a high proportion of RI is used. According to the plot, the adsorption of Cr(VI) is enhanced when the OA content is increased.

The obtained responses may be combined into a single objective function to simultaneously optimise the removals of the different metals. Based on the obtained parameters, the theoretical optimised biosorbent mixture (OAM) composition was obtained: 0.500 g FE, 0.137 g OA and 0.363 g RI.

The analysis of variance (ANOVA) results indicated that the models are significant, and the system can be described using these models. The predicted values were in agreement with the experimental data (Table 3), indicating that the SCMD is a reliable method for determining the optimum composition of the new biosorbent.

According to the model obtained, the expected removal percentages using the optimised mixture were 85.58%, 38.37%, 39.74%, 58.15% and 74.64% for Cu(II), Cr(VI), Ni(II), Zn(II) and Mn(II), respectively. An experiment was conducted using the optimised biosorbent mixture to validate the model with experimental data. The removal levels achieved for Cu(II), Cr(VI), Ni(II), Zn(II) and Mn(II) were 87.10%, 40.90%, 40.73%, 55.61% and 75.69%, respectively. The good agreement between these experimental data and the predicted values along with their standard deviation of less than 5% confirms the validity and adequacy of the model.

### 3.5. Adsorption kinetics and isotherms of the optimised mixture

#### 3.5.1. Adsorption kinetics

When designing a sorption processing system, it is desirable to know the rate at which the metal uptake will occur. Adsorption kinetics provide valuable insights into the mechanism of the reaction and the reaction pathways. Moreover, the time required to complete the process can be estimated.

Fig. 4 shows that metal adsorption increased with time until pseudo-equilibrium was achieved after 180 min. The adsorption process was slower for Cr(VI), probably due to its presence as anion in solution. Some authors have suggested a three-step indirect reduction mechanism for Cr(VI) adsorption in wastes, which would slow the process (Park et al., 2008). That mechanism includes the fixation of anionic chromium to the surface of the biosorbent, the reduction to Cr(III) and, finally, metal complexation with adjacent groups on the surface of the biosorbent. The results of the experiments in which Cr(III) was not detected in the liquid suggest that this three-step mechanism may be involved.

Pseudo-first and pseudo-second order kinetics have been frequently used to describe the experimental data. The models were

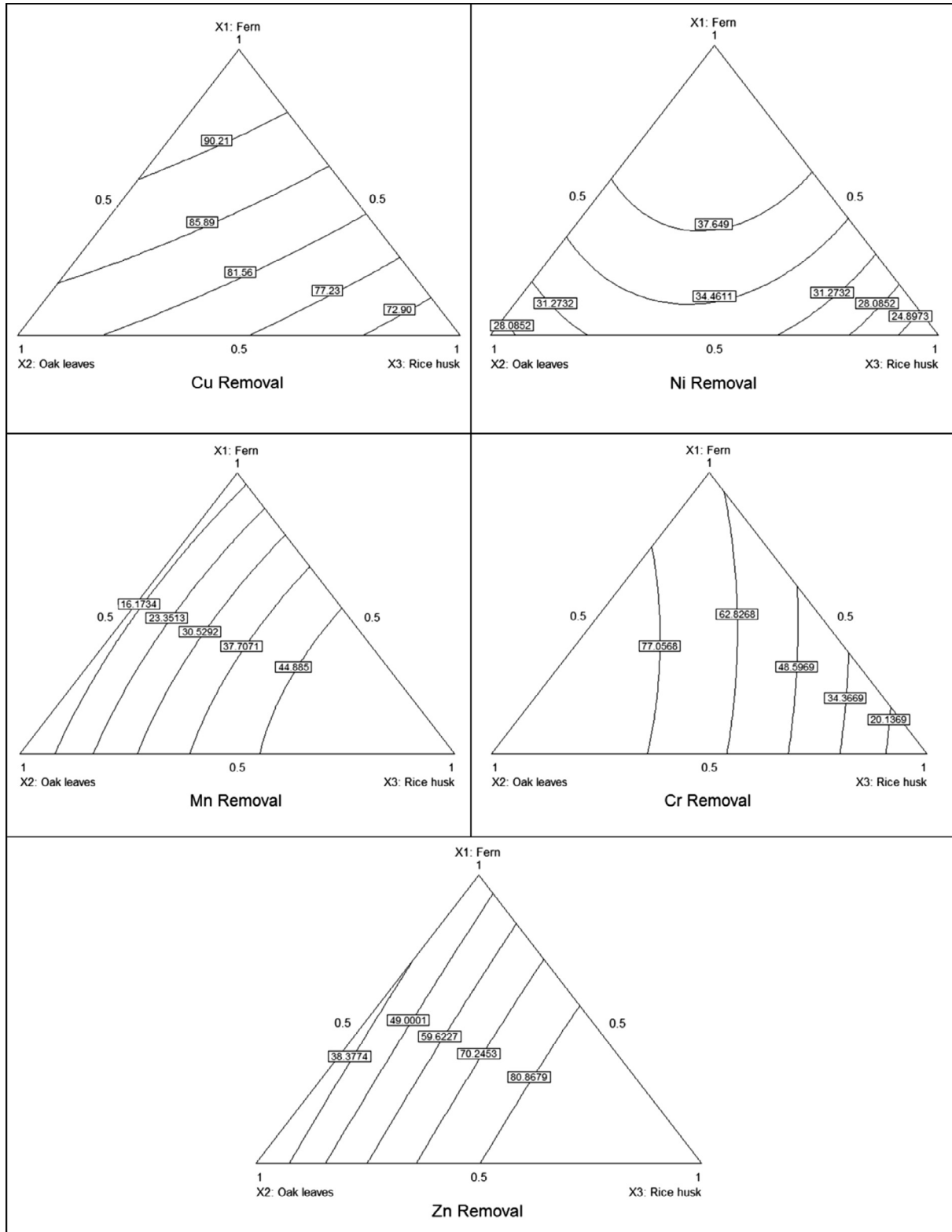


Fig. 3. Triangular-dimensional contour diagrams showing the effect of the different biosorbents on the metal removal.

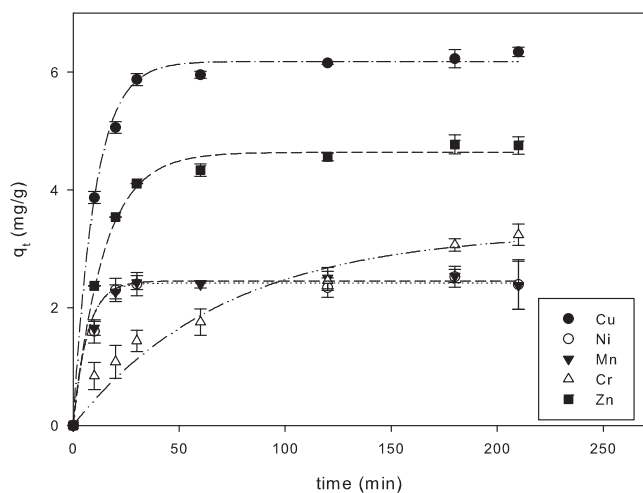
adjusted to the results, and the data were well fit by a pseudo-first order kinetic equation:

$$q_t = q_e(1 - \exp(-k_1 \cdot t)) \quad (4)$$

The kinetic parameters were determined and compared with the experimental values. The calculated values of  $q_e$  agree well with those obtained experimentally. The experimental data are compared with the pseudo-first order model for the different metals in Fig. 4.

**Table 3**  
ANOVA parameters obtained for the SCMD.

Metal	Model	Std. dev.	R <sup>2</sup>	Adjusted R <sup>2</sup>	Predicted R <sup>2</sup>	F value	p-Value Prob > F
Cu	Quadratic	0.7500	0.9948	0.9916	0.9874	308.54	<0.0001
Ni	Quadratic	1.5801	0.9655	0.9439	0.8909	44.71	<0.0001
Mn	Quadratic	2.3820	0.9877	0.9800	0.9591	128.09	<0.0001
Cr	Quadratic	1.5851	0.9980	0.9967	0.9950	786.46	<0.0001
Zn	Quadratic	1.1293	0.9986	0.9978	0.9941	1186.83	<0.0001



**Fig. 4.** Pseudo-first order kinetic fitting to the data obtained under the optimised conditions.

The studied kinetic models cannot identify the adsorption mechanism. Usually, the intraparticle diffusion model is used to identify the adsorption mechanism. The effect of intraparticle diffusion resistance on adsorption can be determined fitting the expression to the data:

$$q_t = k_{id} \cdot t^{0.5} + I \quad (5)$$

where  $k_{id}$  is the intra-particle diffusion rate constant ( $\text{mg}/(\text{g} \cdot \text{min}^{0.5})$ ). Values of  $I$  give information regarding the thickness of boundary layer; i.e., a larger intercept corresponds to a greater boundary layer effect. If the intraparticle diffusion is the only rate-controlling step in the process, the linear plot of  $q_t$  against  $t^{0.5}$  intersects the origin (Rosales et al., 2012). The obtained results showed that the regression is not linear over the whole time range, but can be divided in a few linear regions (Fig. S4). In each of the metals, two linear segments are evident. Therefore, the adsorption process for the studied system followed two stages. The slopes,  $k_{id1}$  and  $k_{id2}$  (Table 4), of the linear segments indicated the rates of the adsorption during the two stages. Therefore, surface adsorption and intraparticle diffusion

contribute to the metal adsorption (Rosales et al., 2012; Chen et al., 2010).

### 3.5.2. Adsorption isotherms

The elucidation of appropriate adsorption equilibrium correlation curves is required to optimise the design of an adsorption system and compare it with different systems for metal removal from polluted waters. Adsorption isotherm models have been widely used to describe the adsorption process and investigate adsorption mechanisms. The equilibrium state has been described by common two-parameter (Langmuir and Freundlich) and three-parameter (Sips and Redlich–Peterson) adsorption isotherm models (Table 5). The three-parameter models reflect the combined features of the Langmuir and Freundlich models. The equilibrium data were analysed using these two- and three-parameter models. Table 5 shows the parameters of the different isotherm equations evaluated for the optimised mixture of biosorbents along and the corresponding linear regression coefficient ( $R^2$ ).

After fitting the experimental data by the different models, the Sips and Redlich isotherms were reduced to the Langmuir expression for all the metals except chromium. The results indicate a better fitting of the experimental data by the Langmuir isotherm.

The favourable Langmuir isotherm adsorption can be expressed in terms of a dimensionless constant separation factor or equilibrium parameter  $R_L$  (Weber and Chakraborty, 1974):

$$R_L = 1/(1 + b_L \cdot C_0) \quad (6)$$

where  $b_L$  is the Langmuir constant, and  $C_0$  is the initial metal concentration. The value of  $R_L$  indicates whether the isotherm is irreversible ( $R_L = 0$ ), favourable ( $0 < R_L < 1$ ), linear ( $R_L = 1$ ) or unfavourable ( $R_L > 1$ ). The equilibrium parameters were calculated for metal adsorption using OAM, and the adsorption is favourable for all of the studied metals.

The adsorption capacity obtained from Langmuir model increases in the order: Ni(II) < Mn(II) < Cr(VI) < Cu(II). This can be attributed to adsorption combined with ion exchange mechanism and, ionic attraction of positively charged metal ions, including chromium positively charged after reduction, and negatively charged carboxylic groups (Krishnani et al., 2008).

**Table 4**  
Obtained parameters for pseudo-first order and intraparticle diffusion model kinetics.

	Parameters	Cu	Ni	Mn	Cr	Zn
Pseudo-first order model	$q_e$	6.1801	2.419	2.4536	3.3039	4.6378
	$k_1$	0.0979	0.1565	0.1474	0.0139	0.0689
	$R^2$	0.9976	0.9962	0.9964	0.9763	0.9946
Intraparticle diffusion model	$k_{id1}$	0.8679	0.3536	0.3418	0.2536	0.7584
	$I_1$	1.1416	0.5535	0.6208	0.0103	0.0251
	$R^2$	0.999	0.8912	0.9366	0.9773	0.9855
	$k_{id2}$	0.0507	0.0061	0.0046	0.2239	0.0668
	$I_2$	5.5826	2.3461	2.4001	0.0189	3.8266
	$R^2$	0.9808	0.0982	0.0636	0.9980	0.9668

**Table 5**  
Fits of the optimised biosorbent mixture metal adsorption data using different isotherm models.

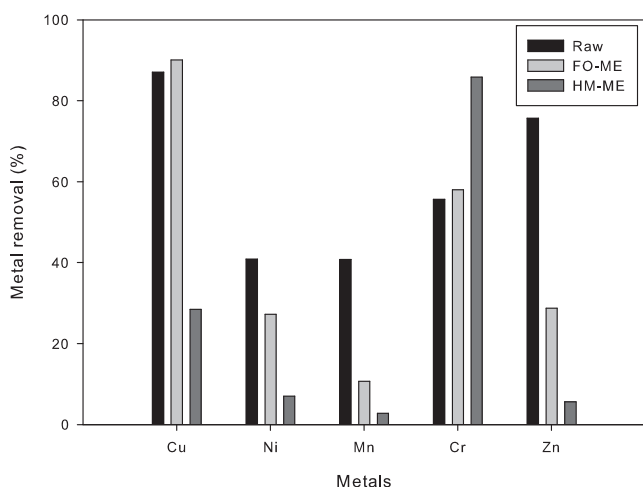
	Parameters	Cu	Ni	Mn	Cr	Zn
Langmuir $q_e = q_{\max} \cdot b_L \cdot C_e / (1 + b_L \cdot C_e)$	$q_{\max}$	32.517	3.901	4.675	23.801	6.823
	$b_L$	0.046	0.194	0.081	0.079	0.599
	$R^2$	0.9786	0.9412	0.9415	0.9579	0.9385
Freundlich $q_e = K_F \cdot C_e^{1/n_F}$	$K_F$	3.365	1.569	1.209	3.341	3.154
	$n_F$	2.400	5.917	4.316	2.687	6.653
	$R^2$	0.9065	0.7103	0.7483	0.9258	0.6950
Sips $q_e = K_S \cdot C_e^{\beta_S} / (1 + a_S \cdot C_e^{\beta_S})$	$K_S$	1.486	0.756	0.382	2.160	4.090
	$\beta_S$	1.000	1.000	1.000	0.8835	1.000
	$a_S$	0.046	0.194	0.087	0.086	0.599
	$R^2$	0.9786	0.9412	0.8715	0.9587	0.9385
Redlich–Peterson $q_e = K_{RP} \cdot C_e / (1 + a_{RP} \cdot C_e^{\beta_{RP}})$	$K_{RP}$	1.499	0.756	0.380	2.386	4.090
	$\beta_{RP}$	1.000	1.000	1.000	0.877	1.000
	$a_{RP}$	0.048	0.194	0.081	0.188	0.600
	$R^2$	0.9786	0.9412	0.9415	0.9660	0.9375

### 3.6. Mechanism of the adsorption process

The effective use of the biosorbent mixture requires understanding the chemical nature of the metal binding process. Therefore, the OAM has been chemically pretreated to inactivate different groups and study the behaviour of the system under those conditions.

When the hydroxyl and phenolic groups of the biosorbent mixture were methylated (FO-ME) using formaldehyde, the mixture's removal capacities for Ni(II), Mn(II) and Zn(II) were significantly reduced. Meanwhile, Cu(II) and Cr(VI) removals were not affected. Therefore, we can conclude that the adsorption of Cu(II) and Cr(VI) is not influenced by the absence of these hydroxyl and phenolic groups.

On the other hand, if the metal binding occurs through interaction with carboxyl groups, chemical modification that reduces the availability of carboxyl groups for metal complexation should cause a marked reduction in their adsorption. Therefore, the optimised biosorbent mixture was pre-treated with HM-ME. This modification esterifies the carboxyl functional groups. The FTIR results showed that the –OH functional group in OAM was replaced by C=O and C–O groups after esterification (Wahi et al., 2013). The esterification significantly reduced the removals of Cu(II), Ni(II), Mn(II) and Zn(II), while the removal of Cr(VI) was significantly increased (Fig. 5). It can be concluded that Cu(II) binding to biomass is mainly related to the availability of carboxylic groups.



**Fig. 5.** Comparison between optimised biosorbent mixture untreated, pretreated with formaldehyde FO-ME and pre-treated with HCl-methanol HM-ME.

**Table 6**  
Recovery percentage of the tested metals for the optimised biosorbent mixture.

	Water (%)	HNO <sub>3</sub> (%)	HCl (%)
Cu	1.71	100.00	100.00
Ni	2.09	100.00	100.00
Mn	2.41	100.00	100.00
Cr	0.17	10.55	5.96
Zn	0.70	97.98	97.81

Moreover, the esterification of OAM also increases the adsorption of anionic metals in solution due to the reduction in the negatively charged groups on biomass surfaces, avoiding charge repulsion.

The other three metals in the synthetic wastewater [Ni(II), Mn(II) and Zn(II)] did not present any clear adsorption preference after the studied pretreatment modifications.

### 3.7. Recovery of metals

The desorption process used to recover the biosorbent is extremely important to assess the biosorbent's potential for reuse. The metal recovery from solution may be based on different mechanisms (precipitation, complexation or ion exchange) (Lezzano et al., 2011).

Different eluents (water, HCl and HNO<sub>3</sub>) were tested to evaluate the recovery of metals in OAM (Table 6). When water was used in the process, there was almost no metal desorption. The use of acidic solutions resulted in a fairly good desorption ability; almost total desorption was achieved for Cu, Ni, Mn and Zn. However, Cr presented almost no desorption, probably due to its presence as an anion as the negative charge is not exchanged with the protons.

The obtained results support the hypothesis of an ion-exchange mechanism being involved in the binding of cations. The results are in accordance with the best fit of the isotherms (Langmuir model), which indicated the dominance of the ion exchange mechanism (Abdolali et al., 2014b). Therefore, HNO<sub>3</sub> and HCl are recommended as desorption agents for the multi-metal mixture.

## 4. Conclusions

The different agroforestry wastes tested appear to have promising potential as biosorbents for metal capture. A significant improvement in removal activity was obtained when a mixture of biosorbents was evaluated and optimised. The selectivity order of OAM in this multi-metal system is Cu > Zn > Cr > Ni > Mn, and the isotherm data are well fit by the Langmuir model. The ion-exchange mechanism is involved in the binding of metals to the biosorbent.



The optimised biosorbent appears to be a promising material for the removal of metals, thus reducing environmental pollution.

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## Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.biortech.2015.01.094>.

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