





Draft Genome Sequences of 12 Shiga Toxin-Producing Escherichia coli Strains Isolated from Dairy Cattle in Portugal

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ABSTRACT Shiga toxin-producing *Escherichia coli* (STEC) is a foodborne pathogen transmitted from animals to humans through contaminated food. Cattle are the main reservoir of STEC, but their genetic diversity is still poorly characterized, especially regarding strains isolated in Portugal. We therefore present the draft genomic sequences of 12 STEC strains isolated from cattle in the north of Portugal.

Shiga toxin-producing *Escherichia coli* (STEC) is able to produce Shiga toxin as its major virulence factor, which is encoded by prophage regions (1). These prophages act as mobile elements and are therefore a driving force in the dissemination of Shiga toxin genes among STEC, other *E. coli* pathotypes, and members of *Enterobacterales* (2). STEC uses the gut of ruminant animals as a natural reservoir and is generally transmitted to humans through the consumption of contaminated foods. From the 500 STEC serotypes known to infect humans, only a few serotypes are responsible for the majority of foodborne cases (3).

In Portugal, the genetic diversity of STEC strains present in cattle has never been evaluated. To increase our knowledge of STEC population epidemiology, the repertoire of STEC genome sequences, and ecology, whole-genome sequencing was performed on 12 strains isolated from dairy cows on a specific farm located in Guilhabreu, Vila do Conde, Portugal.

The genome characteristics and virulence profiles are presented in Table 1. STEC strains were isolated from feces by enrichment with modified tryptone soy broth medium (supplemented with 20 mg liter⁻¹ of novobiocin) and finally cultivated on both MacConkey sorbitol and tryptone bile X-glucuronide agar. Next, typical E. coli colonies were screened for the presence of stx genes by multiplex quantitative PCR (qPCR) according to International Organization for Standardization technical specification 13136:2012. Apart from strains E7N6P4C8A, E7N6P4C8C, and E7N6P4C8F, which are clones isolated from the same animal, the remaining strains originated from different cows/heifers. Strains were grown overnight in brain heart infusion broth at 37°C, and genomic DNA was extracted using the ZR fungal/bacterial DNA miniprep kit (Zymo Research, Freiburg, Germany). The DNA was quantified using a Qubit 3 fluorometer (Invitrogen, Darmstadt, Germany). The sequencing library was prepared with a TruSeq Nano DNA prep kit (Illumina, San Diego, CA, USA) and run on an Illumina MiSeq instrument with 2×251 -bp paired ends using the MiSeq reagent kit v2 (Illumina). The raw data were trimmed with BBDuk (trimming adapters and low-quality reads and discarding reads shorted than 50 bp) (4), normalized (target coverage, 50×; minimum depth, 6×), and assembled with the Geneious Prime 2020 (Biomatters Ltd., New Zealand) de novo assembler with the medium/low-sensitivity settings (5). Read quality and assembly quality were assessed with FastQC v0.11.5 (6) and the Geneious Prime assembler, respectively. Contigs shorter than 500 bp or with coverage less than 8×

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TABLE 1 Sequencing features of STEC strains

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Strain	SKA accession no.	GenBank accession no.	N _{so} (bp)	No. of contigs	No. of raw reads	Genome coverage (x)	Genome size (bp)	No. of CDS ^a	GC content (%)	Serotype	No. of prophages	Shiga toxin gene type(s)	Other virulence genes	gMLST ⁶
E7V3P1C1	SRR12187013	JACBWP0000000000	113,570	82	3,095,354	62	5,459,339	5,376	49.5	O1:H10	14	stx ₁	astA, ehxA, espl, hra, katP, lpfA,	1258
E7V4P1C10	SRR12187014	SRR12187014 JACBWO0000000000	147,484	78	3,120,412	63	5,458,642	5,362	49.4	O1:H10	12	stx ₁	stat, telc, trat astA, ehxA, espl, espP, gad, hra, katP. lpfA. sta1. terC. traT	1258
E7V12P1C4G	SRR12187005	JACBWN0000000000	126,836	105	3,366,256	64	5,352,924	5,449	49.9	0182:H25	25	stx ₁	astA, each, espA, espA, espF, espP, espP, espP, etpD, gad, iss, lpA, nleA, nleB, ompT, tccP, terf, traT	300
E7N3P7C8A	SRR12187006	JACBWM0000000000 130,890	130,890	121	2,751,882	62	5,547,926	5,582	51.1	O150:H2	25	stx ₁ , stx ₂	astA, eae, sin, tun, astA, eae, espA, espA, espI, espJ, espP, gad, iha, iucC, iutA, ipfA, nleB, omnT trCP terC tit traT	306
E7N6P4C8A	SRR12187007	JACBWL000000000	210,932	76	3,759,806	09	5,302,937	5,130	51.3	029:H12	14	stx ₁ , stx ₂	cea, celb, cia, thxA, epeA, gad, hra, iha, lpfA, mchB, mchC, mchF. ompT, terC	155
E7N6P4C8C	SRR12187008	JACBWK000000000	242,239	87	2,707,852	55	5,306,582	5,133	51.0	O29:H12	15	stx ₁ , stx ₂	cea, celb, cia, ehxA, epeA, gad, hra, iha, lpfA, mchB, mchC, mchF. ompT. terC. terC	155
E7N6P4C8F	SRR12187009	SRR12187009 JACBWJ000000000	160,056	62	3,083,908	59	5,302,542	5,126	51.0	O29:H12	15	stx ₁ , stx ₂	cea, celb, cia, ehxA, epeA, gad, hra, iha, lpfA, mchB, mchC, mchF. ompT. terC	155
E7N8P4C1	SRR12187010	JACBW1000000000	238,307	83	2,368,706	51	5,271,944	5,113	51.4	O113:H21	13	stx ₂	cdtB, cea, celb, cia, cib, ehxA, epeA, espP, gad, hra, iha, iss, lpfA, ompT, subA, terC, traT	223
E7N12P2C4	SRR12187011	JACBWH000000000	127,918	44	2,996,158	52	5,811,737	5,937	51.1	O26:H11	22	stx ₁	astA, celb, cif, eae, efa1, ehxA, espA, espB, espF, espJ, espP, fyuA, gad, iha, ip2, iss, iucC, iutA, katP, IpfA, nleA, nleB, nleC, ompT, tcCP, terC, tif, txAB, txT	21
E7N15P4C10	SRR12187012	JACBWG00000000	155,331	72	1,999,722	53	5,182,995	4,968	51.2	O116:H21	11	StX ₂	cba, cdtB, cea, celb, cia, ehxA, epeA, espP, gad, hra, iha, lpfA, ompT, subA, terC, traT	Unknown
E7N16P4C9	SRR12187015	JACBWF000000000	142,475	87	2,079,114	48	5,328,386	5,173	51.1	O29:H12	17	stx ₁ , stx ₂	cea, celb, cia, ehxA, epeA, gad, hra, iha, lpfA, mchB, mchC, mchF, ompT, terC	155
E7N18P5C8G	SRR12187016	E7N18P5C8G SRR12187016 JACBWE000000000	299,977	44	2,479,862	54	5,084,573	4,977	51.9	ONT:H28	10	stx ₁ , stx ₂	cea, cia, cib, cvaC, ehxA, epeA, espP, gad, hra, iha, iss, IpfA, ompT, subA, terC, traT	156

^a CDS, DNA coding sequences. ^b gMLST, genomic multilocus sequence typing.



were removed. The assembly N_{50} values ranged from 111.973 to 299.997 kb. The assembled contigs were screened using VirulenceFinder (7), SerotypeFinder (8), and PHASTER (9) with default parameters for *in silico* typing of *stx* genes and identification of prophages. PHASTER retrieves and classifies a given prophage as incomplete, questionable, or intact on the basis of its genome completeness or potential viability. Draft genomes were annotated using the NCBI Prokaryotic Genome Annotation Pipeline v4.12 (10). The genomic data show a diversity of STEC serotypes and virulence genes, with nine different serotypes detected on a single farm.

Data availability. The assembled genome and sequencing reads were deposited under the BioProject accession number PRJNA643688. The GenBank and Sequence Read Archive (SRA) accession numbers are presented in Table 1.

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