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Polysaccharide from Anacardium occidentale L. tree gum (Policaju) as a coating for Tommy Atkins mangoes

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Policaju-based coatings were applied on "Tommy Atkins" mangoes and the effects of four different treatments on mango shelf-life were evaluated under storage condition at 4°C and 82 % of relative humidity over 28 days. The surface tension of mangoes was found to be 29.04 mN m⁻¹; the dispersive and polar components were 27.57 mN m⁻¹ and 1.47 mN m⁻¹, respectively, and the critical surface tension was 22.7 mN m⁻¹. A significantly lower mass loss was observed in all mangoes treated with Policaju-based coatings. For all applied treatments, no significant variation in the total soluble solids and pH was detected over the experimental storage time. The results show that Policaju-based coatings have a positive effect on the shelf-life extension of mangoes at low storage temperatures (4°C).

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Keywords: Policaju, edible coating, shelf-life, mango

Introduction

Increase of the demand for fresh fruits and vegetables, great losses of quality occurring from the harvesting to the final consumption, and short shelf-life of fruits force the food industry to develop new and better methods of food quality preservation and to extend the shelf-life as well. Besides, all over the world, consumers demand high quality food without chemical preservatives and with extended shelf-life. These factors, taken together, raise the demand for new natural preservatives and antimicrobials (Chien et al., 2007; Lin & Zhao, 2007). Implementation of optimum storage conditions through refrigeration and modified atmosphere or combination of both can be quite effective in maximizing shelf-life and quality of food products. This requires the control of factors such as temperature, relative humidity, gas composition, light, and mechanical/physical stress. However, there is a need for new methods that can be easily applied to smaller amounts of the product at a lower price, while maintaining the preservation efficiency. Edible coatings creating a modified atmosphere surrounding the commodity, with a similar effect to that achieved by

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controlled or modified atmospheric storage conditions, have been considered. Modified atmosphere created by edible coatings protects the food from the moment it is applied until it reaches the final consumer (Durango et al., 2006; Ribeiro et al., 2007). Several researchers have studied the application of coatings on fruits such as apple (Rojas-Graü et al., 2007), strawberries (Ribeiro et al., 2007), mango (Chien et al., 2007; Dang et al., 2008), fresh-cut melon (Oms-Oliu et al., 2008), tomato (Casariego et al., 2008), avocados (Maftoonazad & Ramaswamy, 2005), and litchi fruit (Jiang et al., 2005). Polysaccharide-based coatings are colorless with an oil-free appearance and low caloric content and can be used to increase the shelf-life of fruits, vegetables, shellfish or meat products avoiding or significantly reducing dehydration, oxidative rancidity, and darkening of the surface. Other characteristics that make them attractive are their transport properties, permeability to CO_2 , O_2 , and water vapor, reducing e.g. the mass loss and the microbial spoilage of fruits (Dang et al., 2008). The knowledge of the wettability of coatings is also of particular importance as it is a parameter that defines the ability of a coating to be uniformly distributed on the surface of the fruit (Lin & Zhao, 2007) thus directly affecting its performance as a preservation agent.

Mango is a seasonal fruit that ripens quickly after harvest (between 3 days and 9 days) and, consequently, presents serious restrictions to long distance marketing (Gomez-Lim, 1997). Sensitivity to disease and low temperature as well as the deterioration due to ripening or softening of the fruit, limit its potential in terms of storage, packaging, and transport (Mitra & Baldwin, 1997). However, Shivashankara et al. (2004) showed that lower storage temperatures can be used with Irwin mangoes to improve their shelf-life.

The polyssaccaride obtained from cashew gum (Policaju) is a complex polysaccharide formed by galactose (73.0 %), glucose (11.0 %), arabinose (5.0 %), rhamnose (4.0 %), mannose (1.0 %), and glucuronic acid (6.3 %) with a highly branched galactan framework consisting of chains of $(1\rightarrow3)$ -linked β -D-galacto-pyranosyl units with interspersed β (1 \rightarrow 6) linkages (Paula & Rodrigues, 1995). This nontoxic, hydrophilic, biocompatible, and biodegradable polysaccharide presenting interesting biological activities such as anti-inflammatory and wound healing, has been reported as a potential constituent agent of films (Carneiro-da-Cunha et al., 2009) and suggested as a support for the immobilization of biomolecules (Schirato et al., 2006; Monteiro et al., 2007).

This polysaccharide is a cheap alternative to existing (mostly synthetic) substances and has the advantage of being produced locally, near the harvesting sites of mango. The development of these applications from natural products and its use in the production sites to increase the fruit shelf-life can be an important contribution to the economy of countries like Brazil. In this work, coatings based on the polysaccharide from Anacardium occidentale L. tree gum (Policaju) were evaluated as edible coatings extending the shelflife of mangoes, at a low storage temperature of 4° C, following closely the chemical changes of mangoes over 28 days of storage. Prior to the application of different coatings, the mangoes surface properties were evaluated and the coating composition was optimized in terms of its wettability.

Experimental

"Tommy Atkins" mangoes were purchased from a local producer (Pernambuco, Brazil). Sorbitol 97 % and Tween 80 were obtained from Acros Organics, Belgium. Polysaccharide from A. occidentale L. tree gum (collected from cashew in the South coast of Pernambuco, Brazil) was obtained according to the modified method of Menestrina et al. (1998) and termed Policaju. Briefly, cashew gum (20 g) was crushed and dissolved in distilled water (100 mL) under magnetic stirring (200 min^{-1}) for 2–3 h at room temperature $(25 \,^{\circ}\text{C})$. The solution was filtered through a Vual tissue followed by a new filtration using screen printing cloth (90 thread type), and precipitated with ethanol (1:3). The precipitate (Policaju) was dissolved in distilled water, filtered using screen printing cloth (110 thread type) and finally dried at 30 °C to remove residual ethanol.

Properties of mangoes

As uniform as possible, the mangoes were selected according to the same semi-mature state (color and smell) with similar size, absence of damage and fungal infection and then washed with distilled water. Thin portions of the outer surface (skin) of mangoes were cut with a knife and placed on a glass plate for contact angle measurement.

According to Zisman (1964), in systems with a surface tension lower than 100 mN m⁻¹ (low-energy surfaces), the contact angle formed by a liquid drop on a solid surface will be a linear function of the surface tension of the liquid, $\gamma_{\rm LV}$, (where phase V is air saturated with the vapor of liquid L). The Zisman method is applicable only for low energy surfaces; therefore it is necessary to determine the surface energy of mangoes.

For a pure liquid, if polar $(\gamma_{\rm L}^{\rm p})$ and dispersive $(\gamma_{\rm L}^{\rm d})$ interactions are known, and if θ is the contact angle between that liquid and a solid, the interaction can be described in terms of the reversible work of adhesion, $W_{\rm a}$, as

$$W_{\rm a} = W_{\rm a}^{\rm d} + W_{\rm a}^{\rm p} \Leftrightarrow W_{\rm a} = 2\left(\sqrt{\gamma_{\rm S}^{\rm d}\gamma_{\rm L}^{\rm d}} + \sqrt{\gamma_{\rm S}^{\rm p}\gamma_{\rm L}^{\rm p}}\right) (1)$$

where $\gamma_{\rm S}^{\rm p}$ and $\gamma_{\rm S}^{\rm d}$ are the polar and dispersive contributions of the surface of the studied solid. Rearranging Eq. (1), the following equation is obtained

$$\frac{1+\cos\theta}{2}\frac{\gamma_{\rm L}}{\sqrt{\gamma_{\rm L}^{\rm d}}} = \sqrt{\gamma_{\rm S}^{\rm p}}\sqrt{\frac{\gamma_{\rm L}^{\rm p}}{\gamma_{\rm L}^{\rm d}}} + \sqrt{\gamma_{\rm S}^{\rm d}} \qquad (2)$$

Contact angle determinations of at least three pure compounds: bromonaphthalene (Merck, Germany), formamide (Merck, Germany), and ultra pure water, on the surface of mangoes (fruit skin) combined with the dispersive and polar component values allow cal-

culating both the independent
$$\left(\sqrt{\frac{\gamma_{\rm L}^{\rm p}}{\gamma_{\rm L}^{\rm d}}}\right)$$
 and the de-

pendent
$$\left(\frac{1+\cos\theta}{2}\frac{\gamma_{\rm L}}{\sqrt{\gamma_{\rm L}^{\rm d}}}\right)$$
 variables in Eq. (2).

Surface tension, the dispersive and the polar component were, respectively, 72.10 mN m⁻¹, 19.90 mN m⁻¹, and 52.20 mN m⁻¹ for water, 44.40 mN m⁻¹, 44.40 mN m⁻¹, and 0.00 mN m⁻¹ for bromon-aphtalene, and 56.90 mN m⁻¹, 23.50 mN m⁻¹, and 33.40 mN m⁻¹ for formamide (Busscher et al., 1984).

Estimation of the critical surface tension ($\gamma_{\rm C}$) was performed by extrapolation from Zisman plots (Zisman, 1964). These plots allow calculating the critical surface tension ($\gamma_{\rm C}$) defined as

$$\gamma_{\rm C} = \lim \gamma_{\rm LV} \quad \text{as} \quad \theta \to 0^{\circ}$$
 (3)

Wettability was studied determining the values of the spreading coefficient (W_s) and the works of adhesion (W_a) and cohesion (W_c) . Adhesive forces promote the liquid spreading in a solid surface and cohesive forces promote their contraction. Wetting behavior of the solutions mainly depends on the balance between these forces. Surface tension of the coating solution was measured by the pendant drop method using the Laplace–Young approximation (Song & Springer, 1996).

The contact angle (θ) of a liquid drop on a solid surface is defined by the mechanical equilibrium of the drop under the action of three interfacial tensions: solid-vapor ($\gamma_{\rm SV}$), solid-liquid ($\gamma_{\rm SL}$), and liquid-vapor ($\gamma_{\rm LV}$). The equilibrium spreading coefficient ($W_{\rm s}$) is defined by Eq. (4) (Rulon & Robert, 1993) and its value can only be negative or zero

$$W_{\rm s} = W_{\rm a} - W_{\rm c} = \gamma_{\rm SV} - \gamma_{\rm LV} - \gamma_{\rm SL} \tag{4}$$

where the works of adhesion, $W_{\rm a}$, and cohesion, $W_{\rm c}$, are defined by Eqs. (5) and (6), respectively,

$$W_{\rm a} = \gamma_{\rm LV} + \gamma_{\rm SV} - \gamma_{\rm SL} \tag{5}$$

$$W_{\rm c} = 2\gamma_{\rm LV} \tag{6}$$

The contact angle (θ) and the liquid–vapor surface tension ($\gamma_{\rm LV}$) were measured in a face contact angle meter (OCA 20, Dataphysics, Germany). Samples of the coatings were taken with a 0.75 mm diameter 500 μ L syringe (Hamilton, Switzerland). The contact angle at the mangoes' skin surface was measured by the sessile drop method (Kwok & Neumann, 1999). Measurements were done in less than 30 s. Ten replicates of contact angle and surface tension measurements were obtained at (21.3 ± 0.5) °C. Wettability determinations were performed with Policaju concentrations of 0.015 g mL⁻¹ and 0.03 g mL⁻¹. Tween 80 was used as surfactant to improve the wettability of the solution on the mangoes' surface, five different concentrations were used (0.25 × 10⁻³ g mL⁻¹, 0.5 × 10⁻³ g mL⁻¹, 0.75 × 10⁻³ g mL⁻¹, 1.0 × 10⁻³ g mL⁻¹, and 2.0 × 10⁻³ g mL⁻¹). All coatings contained sorbitol as a plasticizer at the concentration of 0.004 g g⁻¹.

Mangoes treatment and shelf-life analysis

Coating solutions were prepared by dissolving 0.015 g mL⁻¹ or 0.03 g mL⁻¹ Policaju in distilled water under agitation using a magnetic stirrer for 24 h at room temperature (25 °C). Subsequently, sorbitol as a plasticizer and Tween 80 were added to the Policaju solution in concentrations of 0.004 g g⁻¹ and between $0-2 \times 10^{-3}$ g mL⁻¹, respectively.

Washed mangoes were dipped into a 0.5 vol. % sodium hypochlorite solution for 3 min, washed twice with distilled water and left to dry. After drying, they were kept at room temperature ($25 \,^{\circ}$ C, close to the temperature of the wettability tests) and at the relative humidity (RH) of 66.6 % until coating application.

Mangoes were arranged into five groups according to the applied coatings: 0.015 g mL⁻¹ of Policaju (P1.5%), 0.03 g mL⁻¹ of Policaju (P3.0%), 0.015 g mL⁻¹ of Policaju with Tween 80 (P1.5%tw), 0.03 g mL⁻¹ of Policaju with Tween 80 (P3.0%tw), and the control group without coating. All coatings contained sorbitol as a plasticizer at the concentration of 0.004 g g⁻¹. Each group was dipped into the corresponding treatment solution for 2 min, the residual coating being allowed to drip off.

The mangoes were randomly divided into five groups of twelve mangoes each and they were stored at 4 °C and 82 % RH for 28 days. On the 1st, 7th, 15th, and 28th day of storage, three mango samples were peeled off and cut lengthwise to remove the stones. The juice samples were squeezed from the two slices of each mango and collected in sterilized jars in aseptic conditions. Afterwards, the juice was centrifuged at 402 g for 5 min and the flesh total soluble solids (TSS) were determined on the supernatant using a refractometer (Atago Automatic – Master T, Japan). For pH determination, mango flesh was triturated and gauged with a potentiometer (Analyses pH 300 M, Brazil).

The mass loss was evaluated with a semi-precision balance (B-TEC-500, Brazil); all mangoes were indi-

vidually weighed at the beginning of the study and during the storage time. The mass loss was expressed as

$$W_{\rm a} = \Delta W = \frac{m_{\rm i} - m_t}{m_{\rm i}} 100 \%$$
 (7)

where m_i is the initial mass and m_t is the mass at time t. Each physico-chemical analysis was performed in three replicates.

All data for the variables were analyzed and compared using ANOVA (for W_a , W_c , W_s , mass loss, TSS) and the Tukey test (p = 0.05) (for pH) (SigmaStat, trial version, USA) to determine the significance of differences found between the treatments (corresponding to the five groups described here). Regression analysis used for determination of the surface and critical surface tension was performed using SigmaStat (USA).

Results and discussion

The mangoes' skin dispersive component ((27.57 ± 0.01) mN m⁻¹) is higher than the polar component ((1.47 ± 0.01) mN m⁻¹), which shows the ability of this fruit's surface to participate in non-polar interactions; the surface tension of mangoes was (29.04 ± 0.02) mN m⁻¹. The obtained value of the mangoes' critical surface tension was (22.7 ± 0.02) mN m⁻¹, allowing to conclude that the mangoes have a low-energy surface (i.e., below 100 mN m⁻¹) and that the Zisman method is therefore applicable. On the other hand, values of critical surface tension must be lower than those of the surface tension (Dann, 1970), which holds true for all mangoes used in this study.

The wettability of mangoes' skin was studied determining the values of the work of adhesion $(W_{\rm a})$, work of cohesion (W_c) , and the spreading coefficient $(W_{\rm s})$. The presence of Tween 80 decreases the values of $W_{\rm a}$ and $W_{\rm c}$ (Fig. 1) and allows a better (higher) value of $W_{\rm s}$ to be achieved (Fig. 2). The addition of Tween 80 to the solution decreased the cohesion forces substantially, as expressed by $W_{\rm c}$ (Fig. 1), with a statistically significant difference (p < 0.05). Tween 80 reduces the surface tension of the liquid and increases $W_{\rm s}$, improving thus the compatibility of the solution and the fruit's skin surface. Other authors show the ability of Tween 80 to decrease W_c in the fruits improving the wettability of the solutions (Ribeiro et al., 2007). Fig. 2 shows that with the increase of Tween 80 concentration, the value of W_s increases as well reaching an optimal value at the Tween 80 concentration of 2.0×10^{-3} g mL⁻¹, both with Policaju concentrations of 0.015 g mL⁻¹ and 0.03 g mL⁻¹. Similar findings were reported by Carneiro-da-Cunha et al. (2009) using the Tween 80 concentration of 1.0×10^{-3} g mL⁻¹. In our work, a statistically significant difference (p < p)0.05) was also found between these Policaju concentrations and values of $W_{\rm s}$ obtained in the absence of Tween 80.



Fig. 1. Values of the work of adhesion (W_a) and work of cohesion (W_c) of Policaju-based coatings applied on mangoes; Policaju concentrations of 0.015 g mL⁻¹ and 0.03 g mL⁻¹ and different concentrations (c) of Tween 80. Key: (□) W_c P1.5%tw, (■) W_c P3.0%tw, (△) W_a P1.5%tw and (▲) W_a P3.0%tw. Each data point is an average of 10 experiments; error bars show standard deviation.



Fig. 2. Values of the spreading coefficient (Ws) of Policajubased coatings applied on mangoes; Policaju concentrations of 0.015 g mL⁻¹ and 0.03 g mL⁻¹ and different concentrations (c) of Tween 80. Key: (□) Ws P1.5 %tw and (■) Ws P3.0 %tw. Each data point is an average of 10 experiments; error bars show standard deviation.

Four solutions were chosen for further application on mangoes' skin and for the study of the effect of Policaju coatings on mango shelf-life. The solutions were prepared with Policaju concentrations of 0.015 g mL⁻¹ and 0.03 g mL⁻¹, both without and with 0.5×10^{-3} g mL⁻¹ Tween 80. This concentration of Tween 80 was selected because no statistically significant difference was observed between the values of W_s for coatings with the Policaju concentration of 0.015 g mL⁻¹ containing 0.5×10^{-3} g mL⁻¹ or more of Tween 80. For coatings with the Policaju concentration of 0.03 g mL⁻¹, the optimal value of Tween 80 concentration was found to be 2.0×10^{-3} g mL⁻¹,



Fig. 3. Mass loss of "Tommy Atkins" mangoes treated with the following coatings: (○) control, (△) P1.5%, (▲) P1.5%tw, (□) P3.0%, and (■) P3.0%tw. Each data point is an average of 3 experiments; error bars show standard deviation.

corresponding to the $W_{\rm s}$ value of (-23.48 ± 2.09) mN m⁻¹. Though being statistically different from the $W_{\rm s}$ value obtained for the Tween 80 concentration of 0.5 $\times 10^{-3}$ g mL⁻¹ (-27.92 ± 1.55) mN m⁻¹, the gain in $W_{\rm s}$ does not justify a four-fold increase in the concentration of Tween 80. Thereby, the Tween 80 concentration of 0.5×10^{-3} g mL⁻¹ was chosen for both Policaju solutions (0.015 g mL⁻¹ and 0.03 g mL⁻¹) thus reducing the cost of the coating. Coatings with lower values of $W_{\rm s}$ (i.e., those without Tween 80) were also tested on mangoes to evaluate the influence of a decrease of approximately 30 % of the $W_{\rm s}$ value on the shelf-life of the coated mangoes.

Physico-chemical analysis

Change in the mass of the coated fruit has to be monitored during the storage time to determine how effective the coating is as a moisture barrier. It can be considered that mass loss corresponds almost exclusively to water loss since other components that can be lost, such as aromas or flavors, and gaseous products of respiration, are practically undetectable in terms of mass (Olivas & Barbosa-Cánovas, 2005). Fig. 3 shows that on the 28th day of storage there are statistically significant differences (p < 0.05) between the mangoes without coating (control group) and the mangoes with coating. The mangoes with coating presented lower mass loss in comparison with the control group and the difference ranged between 53.2 % and 70.6 %. The best value in terms of mass loss reduction was obtained with the coating containing 0.03 g m L^{-1} Policaju. These differences can be related to the water vapor permeability of the Policaju coating which decreases the water loss of the mango and, consequently, the mass loss. Similar results were obtained by Sothornvit and Rodsamran (2008), showing that

a mango film applied on the whole fruit reduces its mass loss significantly. Assis and Leoni (2003) showed that the mass loss of apples was reduced by 10 % when a chitosan-based coating was used. Furthermore, Ribeiro et al. (2007) presented good results using starch- and carrageenan-based coatings on strawberries and Dang et al. (2008) showed that the mass loss in mangoes covered with a carnauba-based coating was reduced compared with the control group and other coatings.

Differences between the coatings with and without Tween 80 can also be observed in Fig. 3. Coatings with Tween 80 showed a lower mass loss of coated mangoes until the 15th day of storage. However, such difference becomes not statistically significant on the 28th day of storage. As explained by Carneiro-da-Cunha et al. (2009), it is expected that the presence of Tween 80 lowers the WVP of the films, which was observed in the present work.

It is also worthwhile to notice that from the 15th day of storage onwards, the concentration of Policaju seems to be determinant for the observed mass loss, which is higher for 0.015 g mL⁻¹ Policaju films than for 0.03 g mL⁻¹ Policaju films.

Total soluble solids (TSS) are used as an indicator of total sugar content in fruits providing information about the fruit maturation state. TSS is formed by water-soluble compounds of substances like sugars, acids, vitamin C, and some pectins (Oliveira et al., 1999). In the present work, no significant differences (p > 0.05) of the obtained TSS values, ranging between 11.00° Brix and 12.66° Brix, were observed indicating that the coating had no effect on this parameter. These values are in agreement with Brunini et al. (2002) for "Tommy Atkins" mangoes, where the TSS values were also found to be constant during the storage time for all treatments considered. Chien et al. (2007) presented similar results, without variation of TSS, when chitosan-based coatings were used to extend the shelf-life of mangoes. Also Ribeiro et al. (2007) showed that the content of solids in strawberries did not vary significantly during the storage. Sothornvit and Rodsamran (2008) found that the storage temperature was the only factor affecting the TSS in mangoes (Mangifera indica L.) during the ripening. Shivashankara et al. (2004) working with mangoes (cv. Irwin) previously exposed to high electric field treatment and then stored for 20 days and 30 days at 5° C, did not find changes in TSS and concluded that mangoes of cv. Irwin are more suitable for low temperature storage and can be successfully stored for up to 20 days at 5° C without any significant losses in the functional properties and quality attributes.

According to Olivas and Barbosa-Cánovas (2005), water content and water loss in the fruits during the storage should always be considered for the interpretation of soluble solids values. Water loss causes an apparent increase of the concentration of soluble solids

Storage time/days	pH				
	Control	P1.5%	P1.5 %tw	P3.0%	P3.0 %tw
1	4.92 ± 0.36^a	5.02 ± 0.02^a	4.76 ± 0.22^a	5.06 ± 0.15^a	4.66 ± 0.18^a
7	4.93 ± 0.19^{a}	5.10 ± 0.10^{a}	5.11 ± 0.18^{a}	4.89 ± 0.25^{a}	4.62 ± 0.21^{a}
15	5.31 ± 0.39^{a}	5.28 ± 0.15^{a}	5.11 ± 0.04^{a}	4.99 ± 0.41^{a}	4.74 ± 0.31^{a}
28	5.11 ± 0.14^a	5.22 ± 0.26^{a}	5.17 ± 0.04^a	5.04 ± 0.02^a	5.23 ± 0.41^a

Table 1. pH of "Tommy Atkins" mangoes depending on storage time at 4 $\,^{\circ}\mathrm{C}$

a) Equal superscript letters indicate the absence of statistically significant differences (Tukey test p < 0.05).

that may be incorrectly interpreted as a true change in the amount of sugar present in the fruits. Since the mangoes without Policaju-based coatings (control group) showed a higher mass loss, consequently, higher soluble solids concentration occurred in them. As the TSS values were found to be equal for all four treatments and the control group (data not shown), it can be concluded that higher consumption of the previously cited components occurred in the uncoated mangoes during the ripening.

Usually during the storage, an increase of pH occurs as a consequence of the fruits' ripening (Antunes et al., 2003). In general, food decomposition processes caused by hydrolysis, oxidation, or fermentation modify almost always the concentration of hydrogen ion in the food (Oliveira et al., 1999). Therefore, pH is an important parameter in the fruit ripening evaluation. The final organic acid content of the fruit is determined by the net balance of acid synthesis, degradation, utilization and compartmentalization (Müller et al., 1996). Both the citric and the malic acids are the major organic acids present in mangoes and their content decreases during the ripening (Medlicott & Thompson, 1985; Selvaraj et al., 1989; Chen et al., 2009).

Despite the statement above, pH did not change (no statistically significant difference was observed) during the experimental time (Table 1) in all tested groups of mangoes.

The content decrease of O_2 and the consequent increase of CO_2 caused by modified atmosphere decrease the loss of acidity during the storage due to the reduction of enzymatic activity related to respiratory metabolism, increasing the pH of fruits under lower relative humidity (Lima et al., 1996). The fact that the mangoes covered with Policaju-based coatings were stored at 4 °C suggests that such low temperature induced the diminution in the activity of enzymes of the citric acid cycle (Krebs' cycle). Antunes et al. (2003) found similar results when studying the shelf-life of blackberries at the storage temperature of 2 °C. In this way, in our work, the low storage temperature (4 °C) was the cause of pH stability.

Conclusions

Mangoes skins are low-energy surfaces with the

surface tension of (29.04 ± 0.02) mN m⁻¹ (with polar and dispersive components of 1.47 mN m⁻¹ and 27.57 mN m⁻¹, respectively). Critical surface tension for mangoes is (22.7 ± 0.02) mN m⁻¹. Increasing concentrations of Tween 80 resulted in decreasing values of cohesion and adhesion coefficients, thus improving wettability.

Throughout the experiments, mangoes treated with Policaju-based coatings showed lower mass loss in all tested groups, no statistically significant variation was observed for TSS and pH.

Results show that Policaju-based coatings have a positive effect on the shelf-life of mangoes at the storage temperature of 4 °C. From the 15th day of storage onwards, the concentration of Policaju seems to be determinant for the observed mass loss, which is higher for 0.015 g mL⁻¹ Policaju films than for 0.03 g mL⁻¹ Policaju films, independently of the presence of Tween 80.

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