Genetic structure of vineyard-associated Saccharomyces cerevisiae populations revealed by microsatellite analysis

D. Schuller, P. Sampaio, C. Pais and M. Casal

Centro de Biologia, Universidade do Minho, Braga, Portugal



Introduction

temporature and rainfalls, the geographic localization of the vineyard, antifungal applications, the harvest tenthique, agen writing, the vineyard space see and see the vineyard, antifungal applications, the harvest diversity of Saccharomyces sp. strains among the enological fermentative flora. Some strains seem to be which distributed in a glien vilicultural region, can be found in several consecutive years and are also predominant in the immenting flora hypothesizing the occurrence of specific native strains that can be associated to a terroit [1-3].

As a secunity of more immensional products and understandard or many pounds, more as an uncessing upon for specialised wine yeast stains. All present, leading winemakers domand for autochthonous fermenting stains that are able to enhance the oppression of typical sensorial characteristics of wine and ensure the control of the fermentation process, concerning the motto "special yeasts for special traits" [4]. The detailed biogeographical evaluation of fermentative strains is essential for the establishment of adequate selection

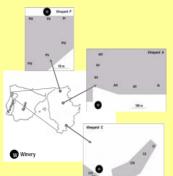
programs.
This is the first systematical, 3-years biogeographical survey of fermentative S. cerevisiae strains using microsaleitiles for the analysis of populational structures and genetic variability in three vineyards of the

Materials and Methods

STAR WASHINGTON TO HIS

The yeas not a non-renneunly playe have too mit was analysed went the must weight was reduced by Yi gif, corresponding to the consumption of about 23 of the sugar content. Must samples were diluted and spread on plates with YPD medium. Thirty randomly selected colonies were collected from each spontaneous fermentation and subjected to further

DNA isolation



Microsatellite amplification

The six trinucleotide microstillie loci described as SchATI, S



Computer assisted data analysis

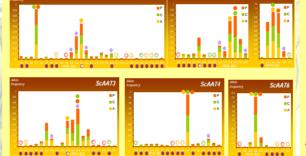
COTI putter assisted Gala ditalysis
A grap of stains with unique microstille profiles (obtained from 30 isolates
per fermentation) was considered the population corresponding to each
sampling site. The pattern and degree of temporal and spatial divergence in the
nuclear microstallities ScAF1 to ScAF6 among subpopulations was
estimated by FST determination over all loci by AMOVA analysis (computed by
the Arlequin software [8]). A similarity matrix of affelic trequencies was
computed by the program NTSYSpc 20 [9], based on the Euclidean distance and
average linkage (UPGMA).

Strains collected

Winery	Year	A1 S _{A1} nples		А6 <mark>А6</mark>	Number of Isolates (*)	Number of genotypes	
А	2001	AT	1	A6	90	11	1
	2002	C1	-	C6	180	34	0 1 2
	2003	C1	-	C6	180	41	1
С	2001	C1	-	Có	240	26	
	2002	P1	+	P6	30	1	
	2003	P1	4	P6	210	35	0 0 2
Р	2001	PI		P6	240	64	000
	2002				150	12	0 2
	2003				300	59	0 2 2

- ollection obtained from this survey comprises 1620 isolates, that were classified in 283 eral samples could not be collected due to a very bad sanitation state of the grapes after heav
- - Number of perennial genotypes (limited to one vinayard)
 Number of annual genotypes (multiple sites of one vineyard)

Allelic frequencies



- respectively.

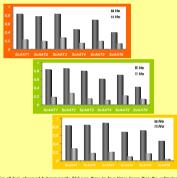
 **A Besides the '4 alleles (51 strains) previously described for ScAATI-ScAATS (3), 52 new alleles were identified in the present study.

 **Some newly described alleles occur with relative high frequency and can be used as indicative alleles for this wine region.

 **All The vast majority of alleles were evenly distributed among 5, cerevisiae populations belonging to vineyard A, C and P, but differ notionized role alleles, which can be considered as integrals', andicative |

 **All Distinct most frequent | • and unique | alleles were found in each of the three populations.

Observed (Ho) and expected (He) heterozygosity

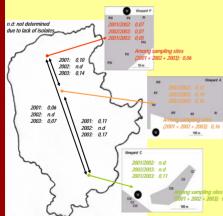


- can be consequence of population substructure.

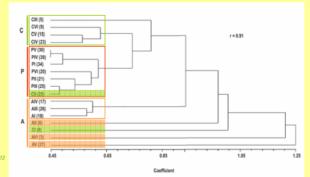
 All Population substructuring can be due to a previously described mechanism called "genome renewal" (10) that has been proposed for natural wine yeast stains, which are mostly diploid and homogogous for the homothalism pine (16490). They can undergo mailing among their propagy calls and thereby change multiple heterographs into completely homograpics cells, leading to gradual replacement of heterograpus diploids:

 Alterograpus difficiencies can also be explained by the presence of null affeles that arise

FST values based on microsatellite data



Relationships among the populations belonging to six sampling points in three vineyards, determined by cluster analysis (UPGMA) based on a Euclidean distance dissimilarity matrix of allelic frequencies. Numbers in parenthesis indicate the number of strains corresponding to unique patterns.



Poplational analysis: Genetic variation of S. cerevisiae populations from vineyards A, C and P.

- 8. AMOVA analysis showed that the S. corevisiae populations from A. C and P were significantly different (Planatom value» cobserved value) 0.0010 in three consecutive years, when populations from different vineyarts were pair wise associated (AC. AP and PIC). FST values were higher for AIC and AP (0.11-0.17 and 0.10-0.10 when compared to PIC (0.06-0.08) corresponding to a moderate (0.65 0.15) to great (0.15 0.25) genetic differentiation.
 Reputations within a vineyard varied in consecutive years, being more variable in A (FST = 0.12 0.19) compared to C (FST = 0.05 0.12) and P (FST = 0.05 0.12).
- When samples were pooled across year-classes within the sampling sites of each vinery, the highest FST value was again obtained for A (0.16) compared to C (0.13) and P (0.06).
- three defined clusters, comprising sampling sites of vineyards C, P and A. Populations within groups C and P are in general more closely related, while S. cerevisiae populations belonging to vineyard A are much more heterogeneous and also more distinct from C and P, which is in accordance with data from FST analysis.
- # The cophenetic correlation factor r was 0.91, indicating that the genetic relationships were no
- Repulation from CII lies within the P-cluster, indicating that genetic differences do not delim
- In Further exceptions from the vineyard specific population structure (sampling sites CIII, All and AVI) may be due to a low number of analyzed strains or to the presence of rare alleles

References

[1] D. Schuller, H. Alves, S. Dequin, M. Casal. 2005. FEMS Microbiol Ecol. 51, 167-177. [2] E. Valero, D. Schuller, B. Cambon, M. Casal, S. Dequin. 2005. FEMS Yeast Res. 5, 959-969 [3] E. Valero, D. Schuller, B. Cambon, M. Casal, S. Dequin. 2005. "Cahiers Techniques",

Lallemand. in press

J Pretorius IS, du Toit M, van Rensburg P (2003). Food Technol Biotechnol 41:3-10. [5] López V., Querol A., Ramón D., and Fernández-Espinar M.T. 2001. Int J Food Microbiol 68:75-81 [6] Pérez, M.A., Gallego, F.J., Martinez, I. and Hidalgo, P. 2001. Lett. Appl. Microbiol. 33, 461-466 [7] Schuller, D., Valero, E., Dequin, S. and Casal, M. 2004. FEMS Microbiol Lett. 231(1) 19-26. [8] Schneider, S., Kueffer, J-M., Roessli, D. and Excoffier, L. 1997. ARLEQUIN version 1.1. Sc

for population genetics data analysis. Geneva, Switzerland
[9] NTSYSpc 2.0, 1997, Exeter Software (http://www.ExeterSoft ww.ExeterSoftw [10] Mortimer, R. K., P. Romano, G. Suzzi, and M. Polsinelli. 1994. Yeast 10:1543-1552.

Conclusions

This poster is available at: http://repositorium.sdum.uminho.pt

Acknowledgements:



Dorit Schuller Centro de Biologia, Campus de Gualtar Universidade do Minho 4710-057 Braga, Portugal Tel: 253 - 60 40 10/17 Fax: 253 – 67 89 80 Mail: dschuller@bio.uminho.pt